

NPAFC
Doc. 1092
Rev. _____

Canadian Highseas Salmon Surveys in 2008-2009

by

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Submitted to the
NORTH PACIFIC ANADROMOUS FISH COMMISSION

by

CANADA

MARCH 2008

THIS PAPER MAY BE CITED IN THE FOLLOWING MANNER:

Trudel, M., J.F.T. Morris, M.E. Thiess, T.B. Zubkowski, and Y. Jung. 2008. Canadian Highseas Salmon Surveys in 2008-2009. NPAFC Doc. 1092. 13 pp. (Available at <http://www.npafc.org>).

INTRODUCTION

The Canadian Program on High Seas Salmon has been conducting integrated epipelagic ecosystem surveys from the west coast of British Columbia to Southeast Alaska since 1998 to assess the effects of ocean conditions and climate change on the distribution, migration, growth and survival of Pacific salmon, and to forecast salmon returns to British Columbia (Beamish et al. 2007; Appendix A). These surveys are usually conducted in late spring-early summer (June-July) and in the fall (October-November). In addition, these surveys have been conducted during winter (February-March) since 2001 to assess the effects of winter conditions on the ecology, bioenergetics, and survival of juvenile salmon (Trudel et al. 2007a).

The working hypothesis of this research is that fast growth enhances the marine survival of salmon, either because fast growing fish quickly reach a size that is sufficiently large to avoid predators (Holtby et al. 1988; Pearcy 1992), or because they accumulate enough energy reserves to better survive their first winter at sea, a period generally considered to be critical in the life cycle of salmon (Beamish and Mahnken 2001). The comparative approach is used to relate changes in salmon growth and bioenergetics to ocean conditions in two contrasting ocean domains: the west coast of Vancouver Island is located at the north end of the California Current System and is affected by upwelling, whereas Southeast Alaska is located in the Alaska Coastal Current and is affected by downwelling (Ware and MacFarlane 1989). In general, sea surface temperature, phytoplankton and zooplankton biomass are higher off the west coast of Vancouver Island (Ware and Thomson 2005).

Three surveys have been planned by the Canadian Program on High Seas Salmon for 2008-2009: a summer survey on June 18 – July 6, 2008, a fall survey on October 7 – November 15, 2008, and a winter survey on February 24 – March 24, 2009. The primary objectives of these surveys will be to (1) collect biological information on Pacific salmon (*Oncorhynchus* spp.) and associated epipelagic fish community, (2) describe the ambient oceanographic conditions, and (3) quantify the biomass of zooplankton and describe zooplankton species community composition in coastal waters of British Columbia and Southeast Alaska.

GENERAL SURVEY INFORMATION

The summer survey will conduct scientific operations primarily on the continental shelf off the west coast of Vancouver Island, in Queen Charlotte Sound, Hecate Strait, Dixon Entrance, Southeast Alaska, and off the west coast of the Queen Charlotte Islands (Table 1; Figure 1). The dates for this survey are currently tentative, as this survey will be conducted using a fishing vessel that will be chartered later this year.

The fall and winter surveys will conduct scientific operations on the continental shelf off the west coast of Vancouver Island, in Queen Charlotte Sound, Hecate Strait, Dixon Entrance, Southeast Alaska, and off the west coast of the Queen Charlotte Islands, as well as the inlets of the west coast of Vancouver Island, central British Columbia, and straits of Southeast Alaska (Table 2-3; Figure 2). These surveys will be conducted using the *CCGS W.E. Ricker*.

FISHING GEAR AND FISHING OPERATION

The research vessel will tow a mid-water trawl, originally manufactured by Cantrawl Nets Ltd., Richmond, BC, and later modified to a model 240 trawl by the fishing crew, for 30 minutes. The trawl has a heavy-duty front end of hexagonal web made from 3/8 in. (9.5 mm) and 5/16 in. (7.9 mm) Tenex rope, and a tapered body made-up of 64 in. (163 cm), 32 in. (81.3 cm), 16 in. (40.6 cm), 8 in. (20.3 cm) and 4 in. (10.2 cm) polypropylene sections, an intermediate section of 3 in. (7.6 cm) polypropylene, and a codend of 1.5 in. (3.8 cm) knotted nylon lined with 0.25 in. mesh (64 mm). The trawl has three 40 m bridles of 5/8 in. (1.6 cm) wire rope per side that are attached with a single hook-up to 5 m Jet doors. Typically, 100-150 m of 1.25 in. (3.2 cm) warp was paid out to tow the trawl at the surface. The trawl is towed at the surface at 5 knots (2.6 m s^{-1}) in good sea conditions, and this typically achieved a mouth opening that was approximately 28 m wide by 16 m deep as measured acoustically by a Scanmar trawl eye mounted on the headrope. In rough weather, the trawl is towed at headrope depths down to 15 m.

The fish samples are sorted by species, enumerated, and measured onboard the ship to characterize the nekton community in epipelagic waters of British Columbia and Southeast Alaska (Brodeur et al. 2006; Orsi et al. 2007). All the juvenile coho and Chinook salmon are systematically scanned for coded-wire tags, irrespective if their adipose fin has been clipped or not, as not all the tagged fish are clipped (Morris et al. 2005). For juvenile salmon, we will also take a skin sample from the operculum using a hole-punch and preserved in 70% ethanol to determine their stock of origin using microsatellite DNA (Beacham et al. 2001, 2005, 2006) and remove calcified-structures (i.e. scales and otoliths) for age determination. Stomach contents of juvenile salmon will also be removed, weighted individually, and pooled by species and tow for dietary analyses (Brodeur et al. 2007). A subsample of the catch is preserved frozen individually at -20°C for various chemical and calorimetric analyses such as stable isotopes, and to examine their stomach contents.

Biological data collected for each salmon will include species common name, fork length, total length, and observed fin clip. It will also include, when available, whole body weight (g wet), sex, stomach content weight (g wet), % water that is based on the ratio of dry to wet whole body weight, coded wire tag number, and pit tag number. Age separation is generally determined based on examination of fork length distributions, that showed non-overlapping size modes for chum, coho, pink, and sockeye salmon (Trudel et al. 2007b). For Chinook salmon, we used a combination of acoded-wire tag recoveries of known-age fish and DNA analyses to establish size-classes to separate juveniles from adults, and life history types (Fisher et al. 2007; Trudel et al. 2007c), as there is considerable overlap among size modes that represent the multiple age groups.

OCEANOGRAPHIC SAMPLING

At oceanographic stations, the scientific crew will (1) conduct CTD (conductivity-temperature-depth) casts, (2) collect seawater samples at 10 m from the surface with a Niskin bottle for nitrate, phosphate, silicate, and salinity, and (3) filter surface seawater on GF/F glass fibre filter disks for chlorophyll a. Nitrate, phosphate, and silicate samples will be collected in acid-washed glass test tubes, whereas the glass fiber disks will be folded and placed in polypropylene scintillation vials. All these samples will be stored frozen. CTD casts will be conducted to 250 m or within 5 m of the bottom with a Seabird

SBE 911+ probe. Several calibration samples from selected CTD casts will be collected over the course of the survey with Niskin bottles at depths where the salinities are stable.

The oceanographic data collected in these surveys will be stored on a database maintained at the Institute of Ocean Sciences (Sidney, British Columbia). The contact procedure to obtain the CTD files is available at:

http://www-sci.pac.dfo-mpo.gc.ca/osap/data/default_e.htm

ZOOPLANKTON SAMPLING

Vertical bongo tows to approximately 150 m or within 10 m of the bottom will be conducted with two 57 cm diameter, 253 µm Nitex nets. One of the nets is equipped with a flowmeter. Zooplankton collected from the flowmeter side will be preserved in 10% formalin and sent to the zooplankton laboratory at the Institute of Ocean Sciences, Fisheries and Oceans Canada (Sidney, BC) for species classification and enumeration. Zooplankton taken from the net without flowmeter will be sorted into four size fractions by successively sieving through 8.0, 1.7, 1.0, and 0.25 mm screens. Each size fraction will then be weighed wet, dried at 60°C for 48 hours, re-weighed, and stored in plastic bags for future stable isotope, bomb calorimetry, and proximate analyses.

The zooplankton data collected in these surveys will be stored on a database maintained at the Institute of Ocean Sciences (Sidney, British Columbia). The contact procedure to obtain detail species records from selected plankton sampling stations is available at:

http://www.pac.dfo-mpo.gc.ca/sci/osap/projects/plankton/zooplanktondatabase_e.htm

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Table 1. Tentative summer survey itinerary for the June 18 – July 6, 2008, DFO High Seas Salmon research survey.

Date	General area of operations
June 18	PBS Nanaimo, loading
June 19-25	West coast of Vancouver Island
June 26	Triangle Island
June 27-28	Hecate Strait
June 29-30	Dixon Entrance
July 1-2	Southeast Alaska
July 2-3	West coast of Queen Charlotte Islands
July 4-5	Travel
July 6	Nanaimo, off loading

Table 2. Tentative fall survey itinerary for the October 7 – November 15, 2008, DFO High Seas Salmon research survey.

Date	General area of operations
October 7	PBS Nanaimo, loading
October 8-18	West coast of Vancouver Island
October 20	Triangle Island
October 21-25	Hecate Strait & Central Coast of British Columbia
October 26-27	Travel
October 28	Nanaimo, off loading
October 29 - November 3	Coast Guard Training Week - No sampling
November 4	PBS Nanaimo, loading
November 5-6	Travel
November 7-8	Dixon Entrance
November 9-12	Southeast Alaska
November 13-14	Travel
November 15	Nanaimo, off loading

Table 3. Tentative winter survey itinerary for the February 24 – March 24, 2009, DFO High Seas Salmon research survey.

Date	General area of operations
February 24	PBS Nanaimo, loading
February 25 - March 5	West coast of Vancouver Island
March 6	Triangle Island
March 7-11	Hecate Strait & Central Coast of British Columbia
March 12-13	Dixon Entrance
March 14-21	Southeast Alaska
March 22-23	Travel
March 24	Nanaimo, off loading

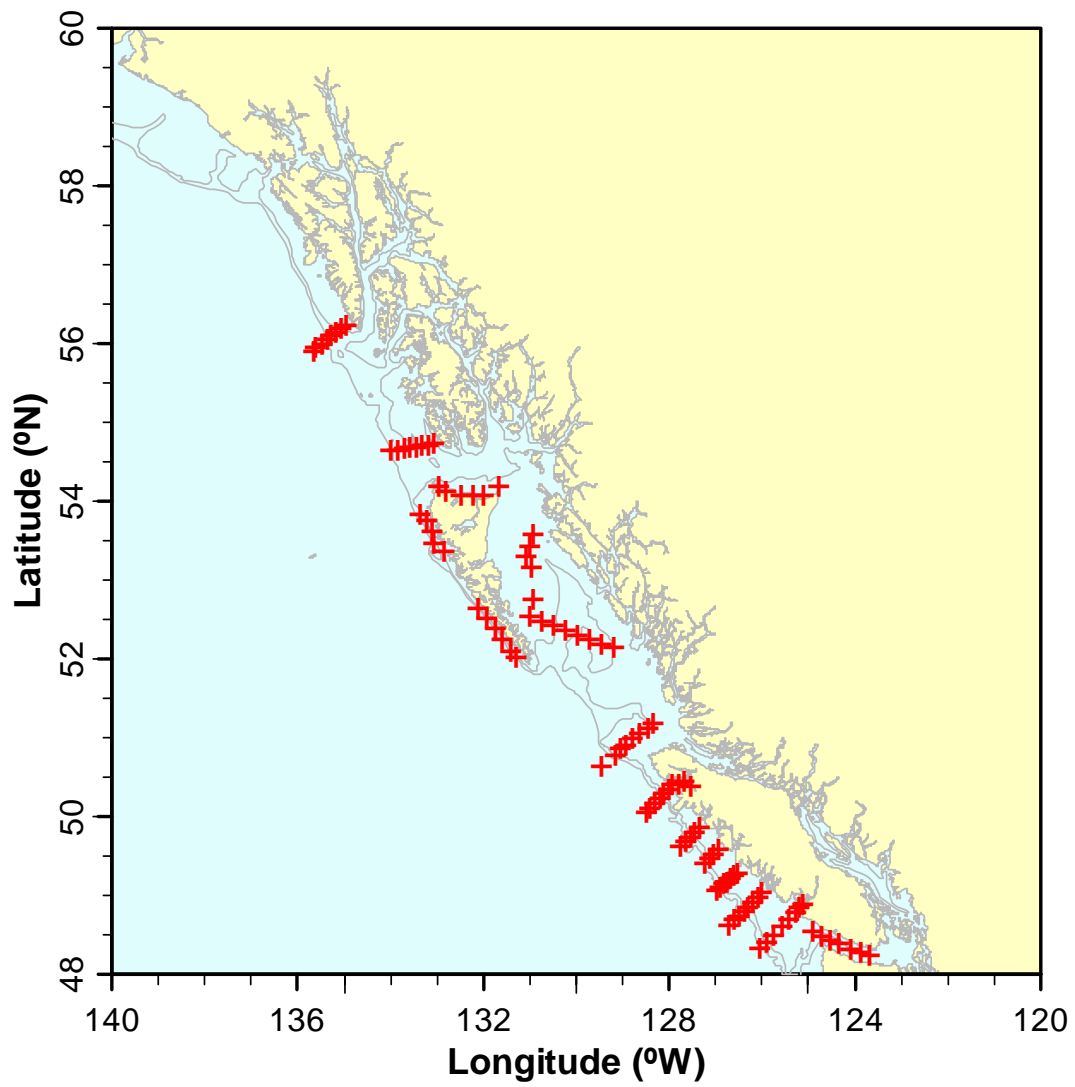


Figure 1. Tentative survey stations for the High Seas Salmon summer survey (June 18 – July 6, 2008).

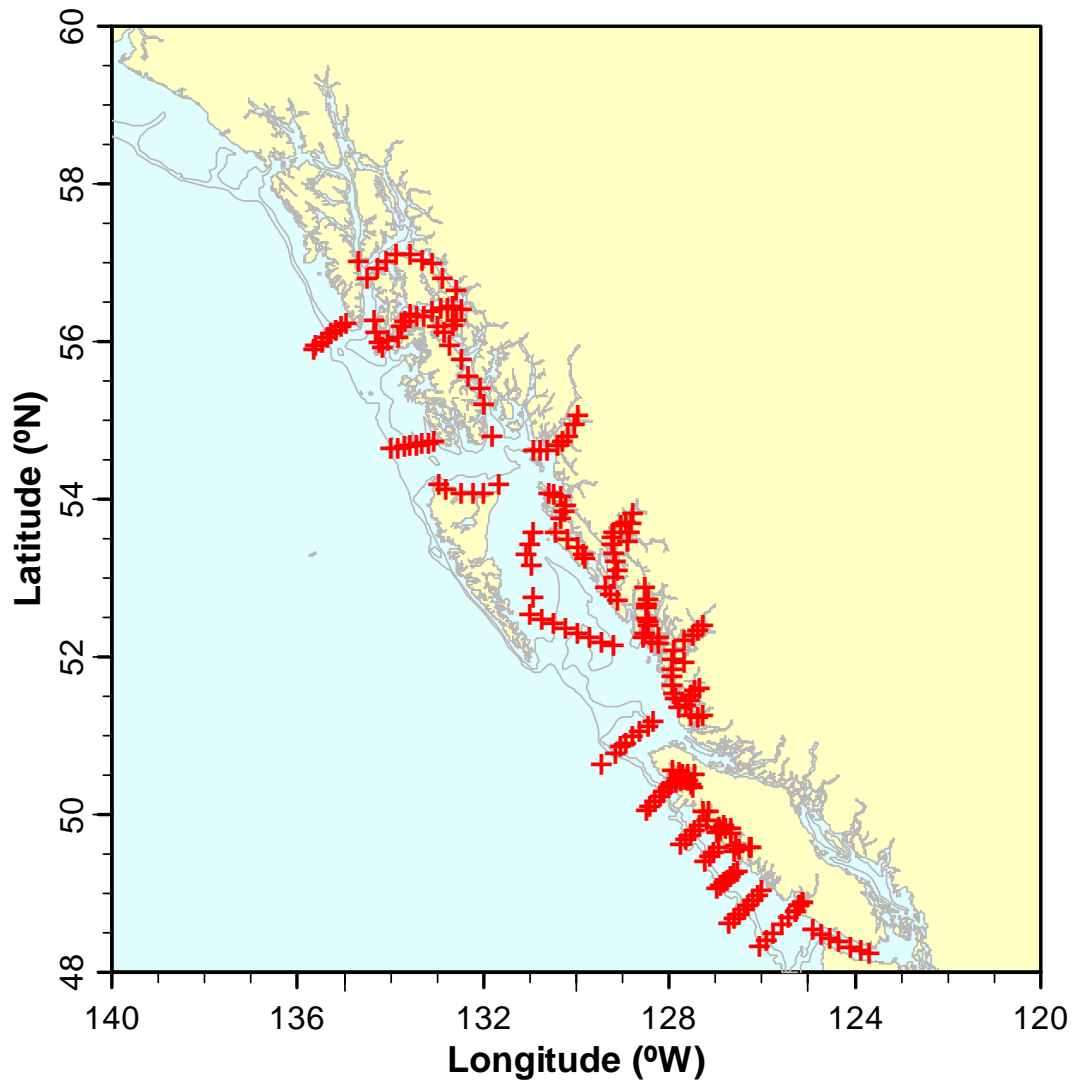


Figure 2. Tentative survey stations for the High Seas Salmon fall (October 7 – November 15, 2008) and winter surveys (February 24 – March 24, 2009).

APPENDIX A. Data reports produced by the Canadian Program on High Seas Salmon.

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